

dimethylcuprate to give the alcohol **14a**. Oxidation followed by immediate reduction after workup afforded the epimer **14b** (87% yield) whose stereochemistry was confirmed by the NOE shown in Scheme II. In order to prepare for the upper "satellite" some straightforward functional group manipulations were required to give **15** and thence **16**. Of special note is the selective hydrolysis of the more-hindered isopropylidene ring engaging the two secondary hydroxyls of **15**.

Elaboration of the upper "satellite" was problematic.³¹ Painstaking investigations educed (a) that a modified³² Corey-Kim procedure³³ was the reagent of choice for selective oxidation of the primary alcohol to the aldehyde **16b** (96%), (b) that the ethylenic acetal **17**, of Sargent and co-workers³⁴ was the preferred version of the Trippett-Bestmann reagent³⁵ for obtaining the 4:1 *Z/E* mixture **16c**, and (c) That both of these geometric isomers upon treatment with methanol and, specifically, Grieco's acid³⁶ led directly to **18** as the only anomer in 86% Yield.

Epoxidation of the olefinic sugar **17** with MCPBA was extremely slow; however, aqueous NBS generated a mixture of bromohydrins, which gave a single epoxide **19**, upon reaction with sodium hydride. The opening of epoxide **19** was effected best by Me_2Mg in THF,³⁷ generated by mixing equimolar amounts of methylolithium and methylmagnesium chloride. The resulting product was methylated to afford the targeted intermediate **1**.

The reactions in Scheme II proceed with very high yields, but the length of the sequence diminishes its attractiveness and hence efforts have been made to provide shorter pathways. We have had encouraging results with respect to the bipyranose **14b** (Scheme III).

By application of Normant's chemistry³⁸ we obtained the vinyllithium derivative **20a** which was converted into the Grignard **20b**. The latter reacted with **8** to give **21** in 60% yield. This

moderate yield is not serious since the only other product is a bromohydrin that is recycled to **8**. Solvolysis of **21** gave the α -anomer **22** predominantly.

Hydroboration of **22** occurred from the *exo* face, giving **14a**, making it possible to obtain **14b** from **8** in five steps instead of the eight shown in Scheme II. Eight more steps are presently required to go from **14b** to **1**, but efforts are under way to reduce these.

Conclusion

There are several features about the new strategy outlined above that are noteworthy:

(a) Each of the seven centers highlighted in the tripyranose **1**, obtained by either route (Schemes II or III), has been created with complete stereo- and/or regioselectivity, and therefore no troublesome separations of isomers are required. This is due to the fact that each ring is a reliable pyranose and hence there are no "off-template" stereocenters! Problem 1 (*vide supra*) is therefore obviated.

(b) Although each center was introduced rationally and was easily monitored, all protons in the entire assembly **1** can be assigned, readily and simply, by [only!] 200-MHz ¹H NMR, as is apparent from the data shown in Scheme II. The easy proof of configuration common to carbohydrate derivatives (see problem 2 above) is therefore observed in **1**.

(c) The folding pattern adopted cuts down upon the number of "external" protecting groups that would be required by a linear array, since **1** (or IX) protects itself. Note also that **1** requires one less protecting group than IX.

(d) Since the oxirane moiety is a synthon for "pyranosidic homologation" (e.g., **8** → **12**), an iterative procedure should be applicable to either **13** or **19**.

(e) Retroanalysis of many other propionate-derived entities, e.g., the ansa chains of streptovaricin and maytansine, according to the five principles outlined above indicate other opportunities for the pyranosidic homologation approach.

The two tasks remaining for conversion of **1** into **2** are deconvoluting the tricycle and adding the C-24 CH_3 . Procedures for carrying these out are being developed.

Registry No. **1**, 88392-88-5; **8**, 33208-47-8; **9**, 88412-16-2; **10**, 88392-78-3; **11**, 88392-79-4; **12a**, 88392-80-7; **12b**, 88412-17-3; **13**, 88412-18-4; **14a**, 88392-81-8; **14b**, 88424-60-6; **15**, 88392-82-9; **16a**, 88392-83-0; **16b**, 88392-84-1; **16c**, 88392-85-2; **17**, 78950-65-9; **18**, 88392-86-3; **19**, 88392-87-4; **20a**, 88392-89-6; **21**, 88412-19-5; **22**, 88392-90-9; rifmaycin S, 13553-79-2; levoglucosan, 498-07-7.

Supplementary Material Available: ¹H NMR data for compounds **1**, **10**, **12**, **13**, **14a**, **14b**, **16b**, **16c**, **18**, **19**, **21**, and **22** (6 pages). Ordering information in given on any current masthead page.

Robustadials A and B from *Eucalyptus robusta*

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Abstract: The structures of two new compounds, robustadial A and B, isolated from the active fraction of the antimalarial extract of *Eucalyptus robusta* leaves, a plant used in Chinese herbal medicine to treat malaria, have been determined as **2** and **3**. The structures were resolved by using spectroscopic methods, especially two-dimensional NMR and difference nuclear Overhauser enhancement techniques. The monoterpene moiety containing the spiro linkage has not been encountered previously.

The leaves of *Eucalyptus robusta* Sm. (Myrtaceae) are used in Chinese herbal medicine for the treatment of dysentery, malaria,

and other bacterial diseases. The benzene-soluble fraction of the crude 95% ethanol extract of the leaves showed significant inhibition against *Plasmodium berghei*, a malaria-inducing protozoan. From this fraction, an active compound, robustadial A (**1**) was isolated.¹

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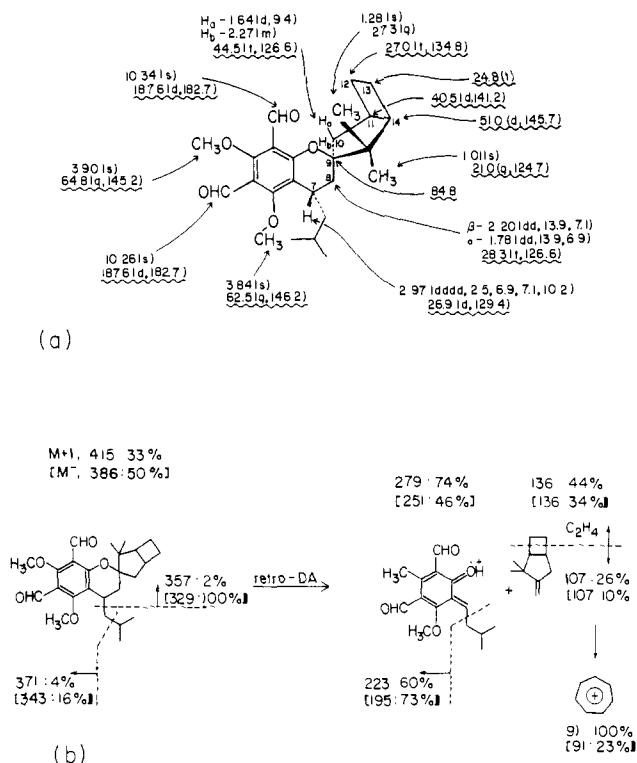
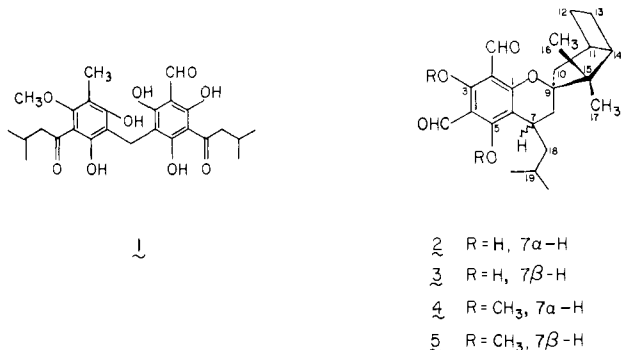


Figure 1. (a) ¹H NMR (250 MHz CDCl₃) and ¹³C NMR (62.9 MHz CDCl₃; underlined) of **5**. *J* values in ¹³C NMR data denote ¹J_{13C-H}; the *J* value for C-13 could not be measured due to severe overlap in undercoupled spectrum. See Figure 2 for the ¹H NMR data not shown here. (b) Desorption chemical ionization (NH₃ carrier gas) MS of **5** and electron-impact MS [in square brackets] of **3**; peak heights are shown in % relative to base peak.

A second fraction contained several compounds that possessed considerably greater activity than **1**.² We now report the isolation and structure of two of these compounds, robustial A (**2**) and robustial B (**3**). The benzene-soluble fraction was partitioned



with 2% NaOH, and the aqueous layer subsequently was acidified and partitioned with benzene. The active benzene fraction was chromatographed (silica gel, petroleum ether) to give **1** and an oil, which was further chromatographed on silica gel (petroleum ether/EtOAc, 50/1) and then on a Lobar pre-packed reverse-phase column (RP-8, MeOH/CHCl₃, 95/5) to yield a mixture of two isomeric compounds, **2** and **3**, which could not be readily separated without derivatization.³ As preliminary spectral data of the mixture suggested that **2** and **3** were phenolic: ¹H NMR (CDCl₃)

(1) Qin, G. W.; Chen, H. C.; Wang, H. C.; Qian, M. K. *Acta Chim. Sin.* **1981**, *39*, 83.

(2) Qin, G. W.; Gu, X. M.; Lu, B. F.; Xu, R. S., unpublished results.

(3) Compounds **2** and **3** were finally separated by using reverse-phase HPLC (JASCO, YMC-gel, ODS S-5, 150 × 4.6 mm, CH₃CN/H₂O, 85/15) after an exhaustive examination of numerous conditions of columns and solvent systems.

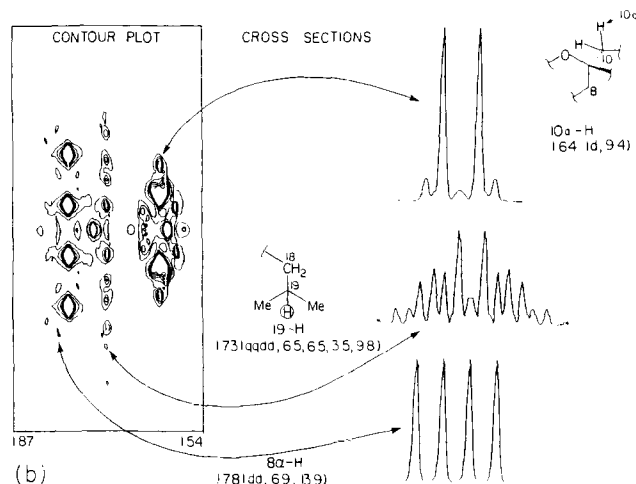
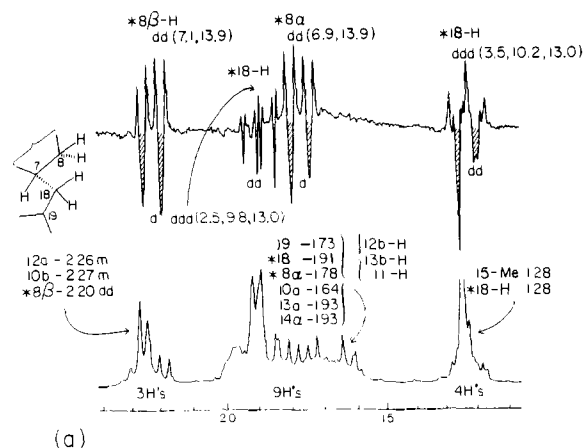


Figure 2. (a) Difference decoupling spectrum of **5** (CDCl₃) obtained by subtracting the spectrum of **5** with decoupling of 7-H (2.97) from the undecoupled spectrum. The asterisked protons are those which underwent decoupling. The δ 's of the 12b-, 13b-, and 11-protons could not be read accurately. (b) *J*-Resolved 2D NMR spectrum of **5** (CDCl₃) for determination of coupling partners of 7-H.

δ 13.52, 13.60; IR (NaCl) 3000 cm⁻¹ (broad, OH), 1635 cm⁻¹ (chelated aldehyde); the mixture was methylated (CH₃I/K₂CO₃/acetone) to yield the respective dimethyl ethers **4** and **5** which were readily separated on HPLC (Whatman Porasil, CH₂Cl₂/0.3% *i*-PrOH). The spectral properties of **4** and **5** were nearly identical. **5**: UV(MeOH) 261 nm (ϵ 11 500), 280 (9200), 320 (2880); CD (MeOH) 227 nm ($\Delta\epsilon$ -2.54), 257 (-2.50), 292 (+1.15), 330 (+0.47), 354 (+0.19); FTIR (NaCl) 1686 cm⁻¹ (CHO); HRMS *m/e* 414.2416 (M⁺, C₂₅H₃₄O₅; calcd, 414.2406). **4**: CD(MeOH) 235 nm ($\Delta\epsilon$ +1.96), 259 (+3.86), 275 (+2.29), 320 (+1.81), 354 (-0.59); MS (DCI) 415 (M + 1, 33%).

The UV and IR spectral data for **5** as well as the ¹³C NMR and ¹H NMR spectra (CDCl₃) suggested a fully substituted aromatic acetogenin moiety bearing two aldehydes and three ether linkages, two of which are methoxyl groups. The remaining benzylic position is occupied by a tertiary carbon bearing an isobutyl side chain as shown by NMR and MS (Figure 1).⁴ The acetogenic chromophore plus the isobutyl side chain account for C₁₅H₁₈O₅ and six units of unsaturation leaving a C₁₀H₁₆ moiety and three units of unsaturation which must include two terpeneoid-Me groups, ¹H NMR (CDCl₃) δ 1.01 (s) and 1.28 (s). The mass spectra of **4** and **5** show a retro-Diels-Alder cleavage as their major fragmentation pathway, yielding peaks at *m/e* 279 (74%, C₁₅H₁₉O₅) and 136 (44%, C₁₀H₁₆), requiring a six-membered ring fused to the aromatic nucleus. Furthermore, the third ether bond links the aromatic ring to a quaternary carbon (δ 84.8 s). With

(4) The MS (DCI) of **2** and **3** both show the loss of a C₄H₈ moiety as the major fragmentation pathway. This loss is ascribed to cleavage of the isobutyl side chain at the benzylic position.

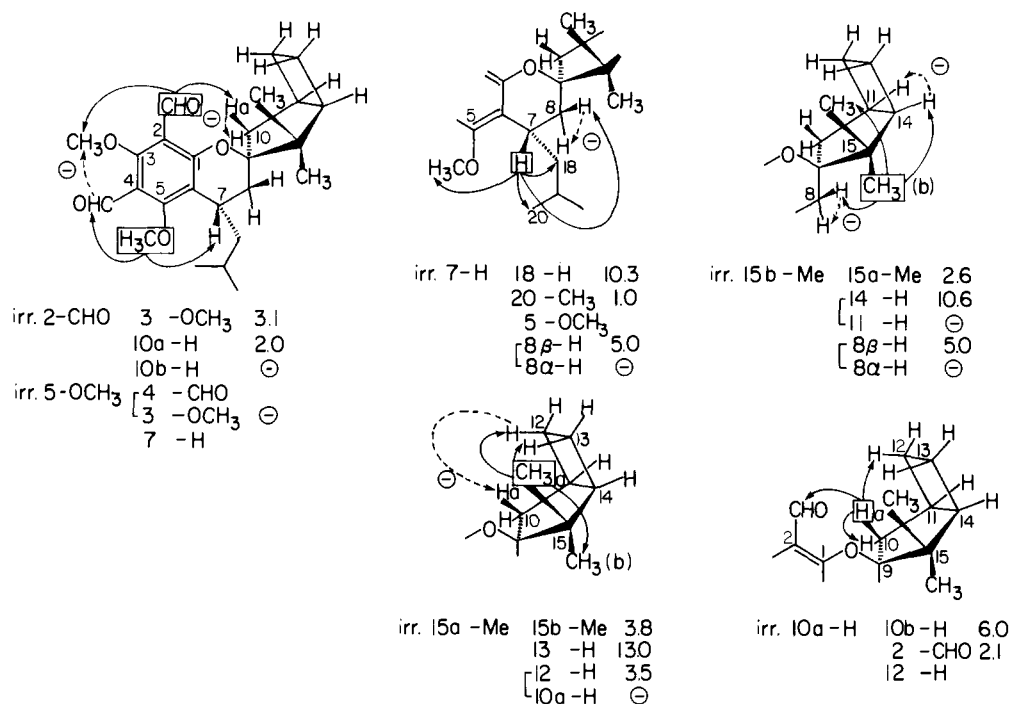


Figure 3. Difference NOE (%) of **5**, 250 MHz CDCl₃. The dashed arrows denote negative NOE's. The % increment of some positive NOE's could not be measured accurately due to overlap of methyl singlet.

the requirement of a six-membered B ring, the remaining six non-methyl carbons must form a bicyclic structure to account for the final two units of unsaturation. The absence of high-field signals in the ¹³C and ¹H NMR spectra rules out a cyclopropyl unit.

The nine overlapping proton signals in the 1.5–2.0 ppm region were clarified by the techniques of difference decoupling (Figure 2a) and *J*-resolved 2D NMR (Figure 2b). Irradiation of 7-H (Figure 2a) decoupled four protons, two each attached to C-8 and C-18. Therefore C-7 (benzylic) is vicinal to two methylenes, one of which belongs to the isobutyl side chain; the qqdd signal of the isobutyl 19-H is resolved in Figure 2b. The methylene group at C-8 requires that the remaining six non-methyl carbons form a bicyclo[3.2.0]heptane unit linked to the B ring via a quaternary spirocenter at C-9. The appearance of a peak in the high-resolution mass spectrum at *m/e* 28 (C₂H₄) as well as the large ¹J_{C-H} coupling constants,⁵ 51.0 (d, 145.7), 40.5 (d, 141.2), 27.0 (t, 134.8), and 24.8 (t) (Figure 1a) confirms the presence of a bimethylene bridge in a cyclobutyl moiety.⁶ Thus the *gem*-dimethyl and the spiro center are both located on the cyclopentyl ring. This spiro center is not vicinal to a bridgehead carbon of the bicyclo[3.2.0]heptyl moiety, as shown by the multiplicities of the 10-H's (Figure 1a) as well as the nuclear Overhauser enhancements (NOE) (Figure 3).

(5) Wehrli, F. W.; Wirthlin, T. "Interpretation of Carbon-13 NMR Spectra"; Heyden: Philadelphia, 1978; pp 50–52.

(6) Carbon assignments are based on one- and three-bond selective decoupling and selective population transfer experiments. Pachler, K. G. R.; Wessels, P. L. *J. Magn. Reson.* **1973**, *12*, 337.

The relative stereochemistry of the cyclobutyl fusion, the location of the *gem*-dimethyl at C-15 rather than C-10, and the relative stereochemistry at the C-7 position (as the β-epimer) were all confirmed by appropriate NOE's (Figure 3). That dimethylrobustadial A (**4**) is the C-7 α-epimer of **5** was suggested by the close similarity of the ¹H NMR and ¹³C NMR spectra except for the downfield shift of its C-8 peak at 31.0 ppm. The C-7 stereochemistry was confirmed by a 5% NOE enhancement observed in **4** for the 10-Hb proton upon irradiation of 7α-H. All other NOE's observed in the spectra of **4** were in accord with the assigned structure.

Robustadial A and B are structurally related to the euglobals, potent granulation inhibitors isolated from *Eucalyptus globulus*.⁷ Further studies on the absolute stereochemistry of **2** and **3** as well as their biological activities are in progress.

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Registry No. **1**, 88130-98-7; **2**, 88130-99-8; **3**, 88197-30-2; **4**, 88131-00-4; **5**, 88197-31-3.

(7) (a) Sawada, T.; Kozuka, M.; Komiya, T.; Amano, T.; Goto, M. *Chem. Pharm. Bull.* **1980**, *28*, 2546. (b) Kozuka, M.; Sawada, T.; Kasahara, F.; Mizuta, E.; Amano, T.; Komiya, T.; Goto, M. *Ibid.* **1982**, *30*, 1952. (c) Kozuka, M.; Sawada, T.; Mizuta, E.; Kasahara, F.; Amano, T.; Komiya, T.; Goto, M. *Ibid.* **1982**, *30*, 1964.